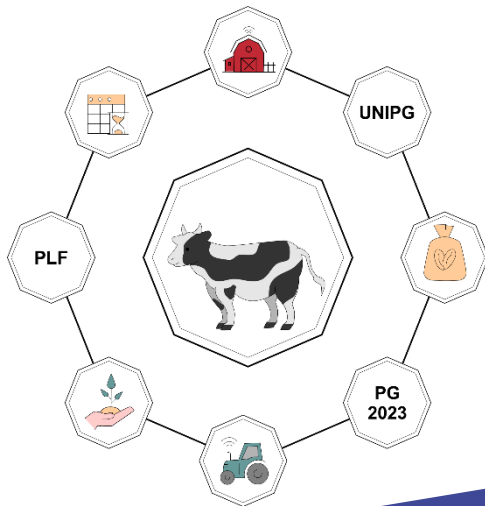


SEMINAR: Infrared Thermography



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*Dp. Agronomy. Animal Production Area.
ETSIA. University of Seville (Spain)*



Índex

1. What is Infrared Thermography?
2. Applications in Animal Production.
3. How do we use it?
4. Practical exercises.

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What is Infrared Thermography?

It refers to the **recording of temperature** using **infrared radiation** emitted from a **body surface**, forming an image called a **thermogram**.

Thermograms are **visual representations** of the amount of **infrared energy emitted, transmitted, and reflected** by an object.

Thermographic Cameras or **Thermal Imaging Cameras** detect radiation in the infrared range of the electromagnetic spectrum (*9,000-14,000 nm or 9-14 microns*) and **produce thermograms** based on the **amount of heat dissipated** at the surface.



The **amount of radiation emitted** by an object **increases with temperature** and depends on their natural heat production (*homeothermic animals versus poikilothermic animals*)

Thermography **allows** you to see **temperature variations**.

What is Infrared Thermography?

Infrared Thermography or **IRT**, measures **infrared emitted rays** in objects, while the usual cameras measure the **light falling** on objects. The **amount of heat** produced **varies** according to the **animal**, the **region** from the body and their **physiological conditions**.

The **measurement** of the **IRT** of an object/individual **depends** on:

- ❖ **EMISSIVITY**: Represents the **material's ability to emit thermal radiation**. Range: 0.00 (*no light output*) to 1.00 (*all light output*).
- ❖ **TEMPERATURE and HUMIDITY**: They **change Emissivity** and therefore, the thermal radiation transmitted.

OTHER FACTORS AFFECTING IRT ASSESSMENT

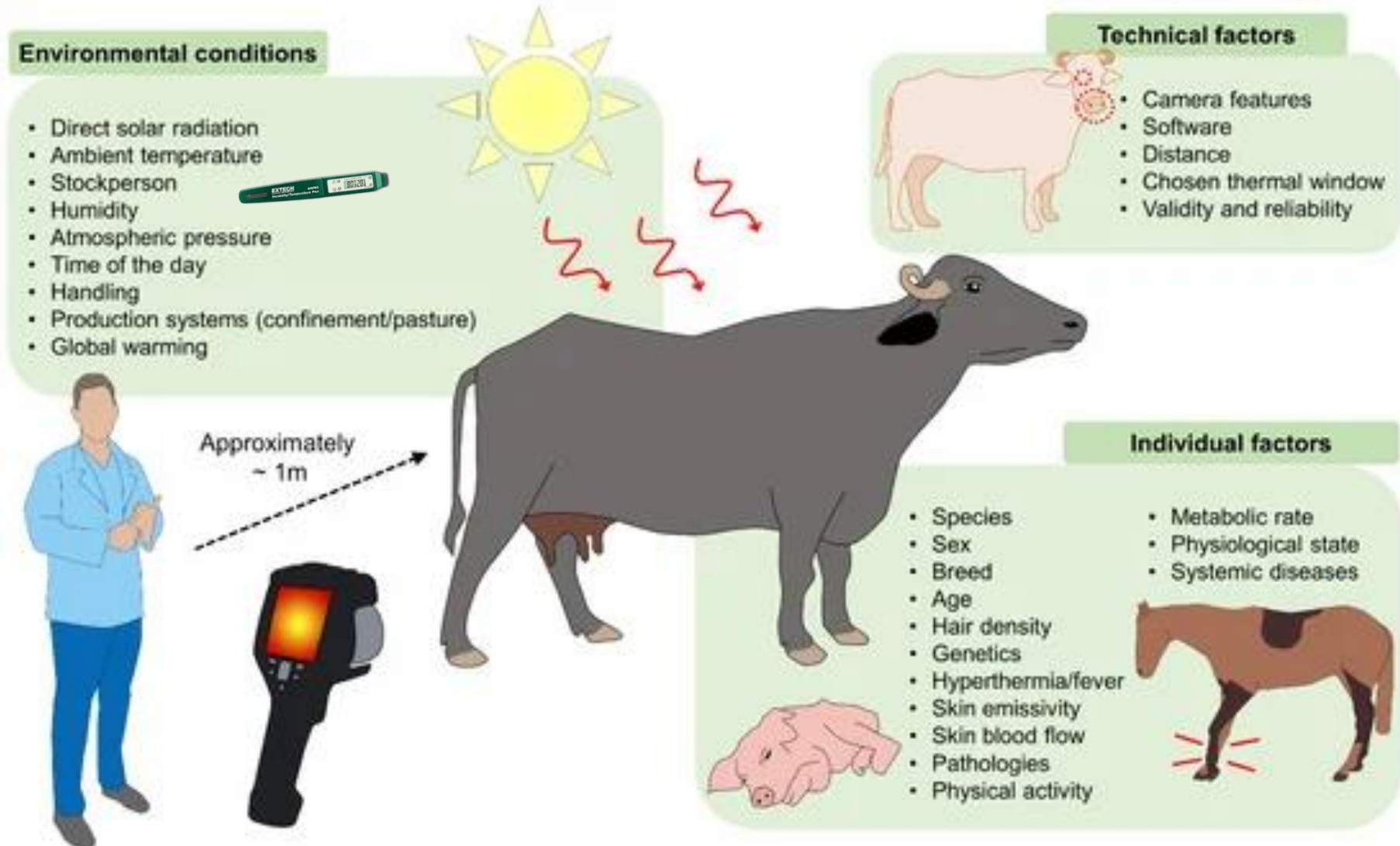


Figure obtained from Mota-Rojas, et al. (2021) *Animals*, 11, 2247.

- An **Infrared Camera** senses infrared radiation, enabling us to **visualize** the thermal world.
- A **thermal imaging camera** consists of **5 components**:
 1. An **optic** system
 2. An **infrared detector**
 3. An **amplifier**
 4. A **signal processing** system
 5. A display **monitor**.

What is Infrared Thermography?

- Some examples of IRT cameras:



<https://es.trotec.com/shop/>



<https://www.flir.es/>



<https://www.fluke.com/es-es>



<https://www.iciscientific.com/>

Índex

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GENERAL THERMAL IMAGING APPLICATIONS

- ▶ INDUSTRIAL APPLICATIONS
- ▶ MEDICINE APPLICATIONS
- ▶ SECURITY APPLICATIONS
- ▶ BUILDING CONSTRUCTIONS
- ▶ NIGHT VISION



ANIMAL PRODUCTION APPLICATIONS

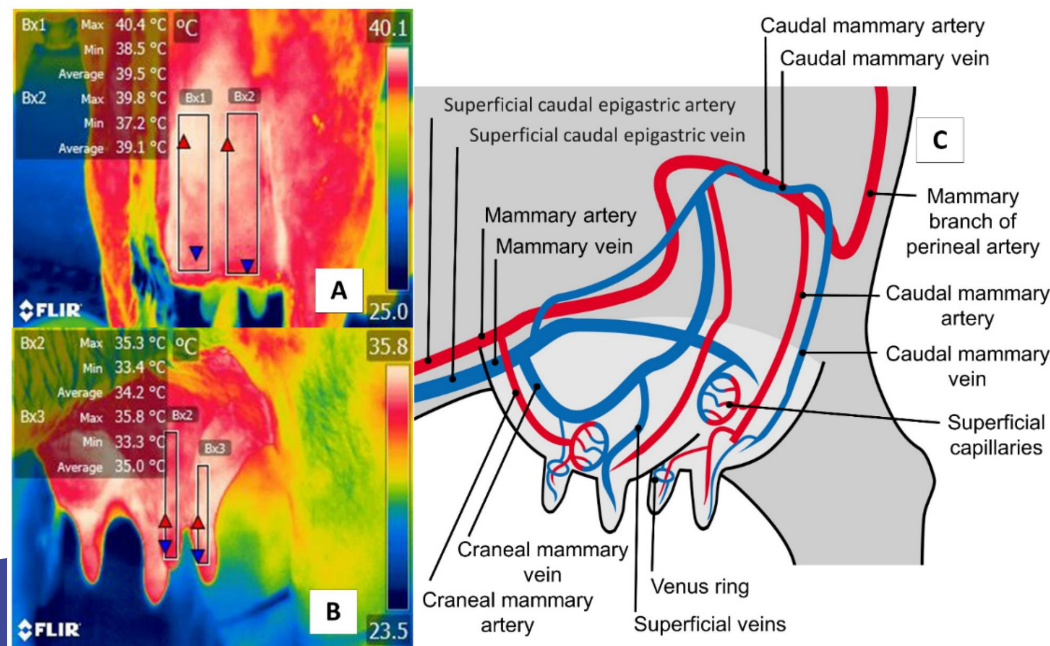
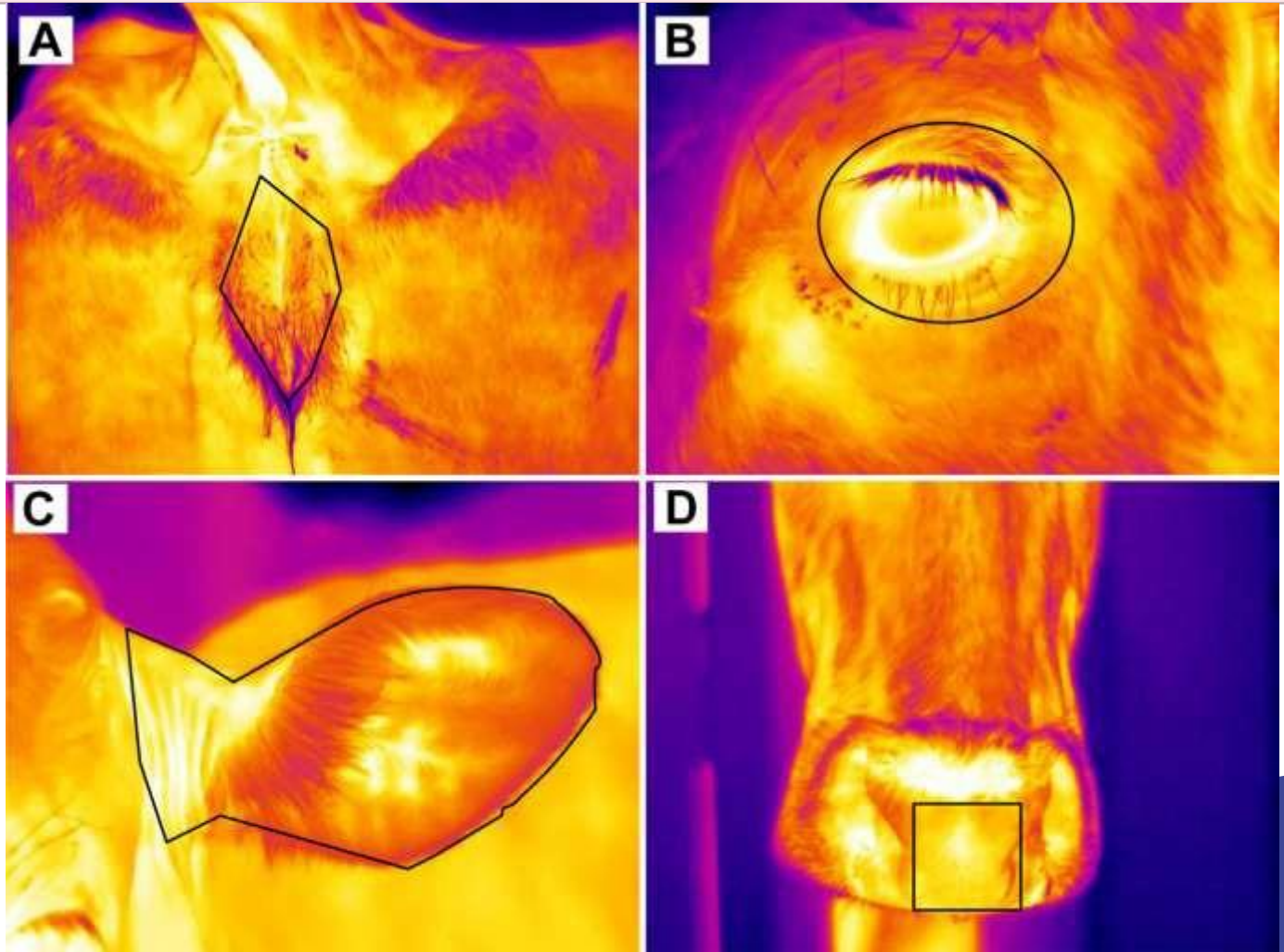
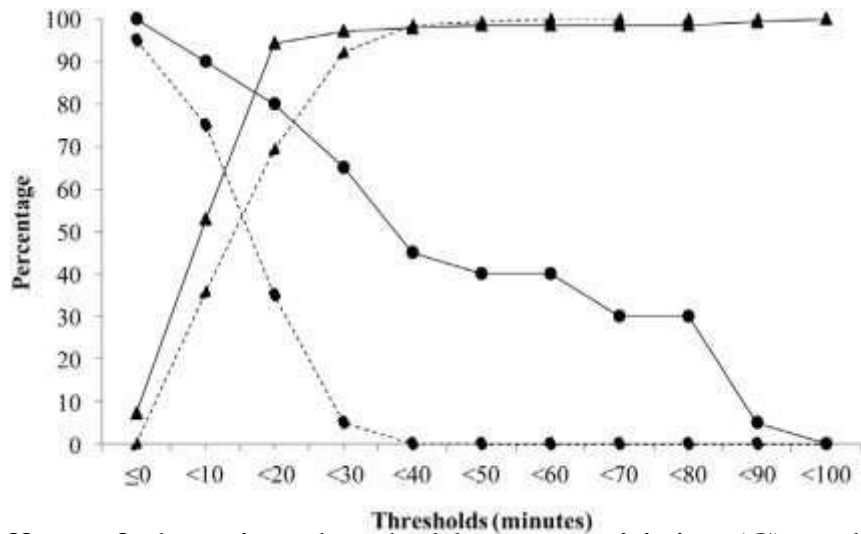


Figure obtained from Mota-Rojas, et al. (2021) *Animals*, 11, 2247.

Body areas	n	Mean
Vaginal temperature	1142	38.5
Vulva	1140	35.0
Left eye	1115	36.3
Right eye	1113	36.2
Left ear	1114	34.8
Right ear	1114	34.6
Muzzle	1112	32.8
Daily activity level	14990	835.2
Daily rumination level (min)	14990	502.0
Daily milk production (kg)	665	28.71
Milk protein (%)	665	2.74
Milk fat (%)	665	3.97

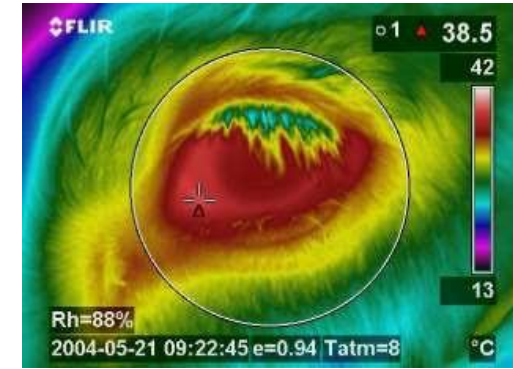


Hand-drawn area of vulva (A), eye (B), ear (C), and muzzle (D), which defined the temperature data area used by the support software.

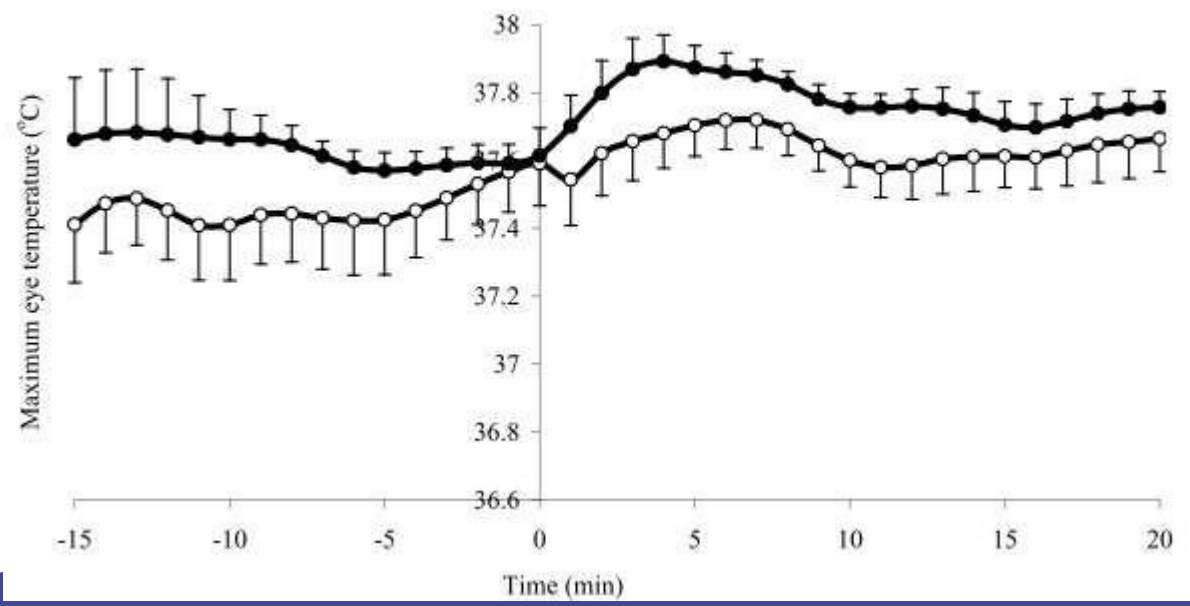
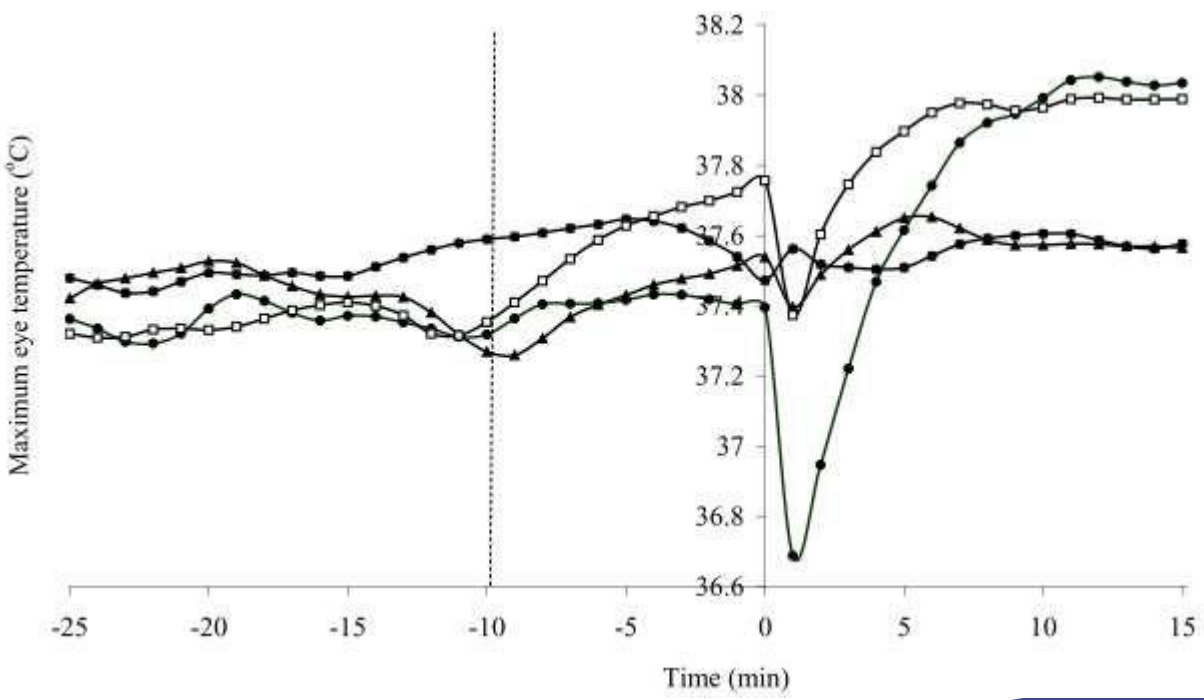
Effect of changing thresholds on sensitivity (C) and specificity (:) for SCR tags measuring activity (solid line) and rumination level (dashed line).

Stewart, M. (2008) Non-invasive measurement of **stress and pain** in cattle using infrared thermography. PhD thesis.
 Massey university, Palmerston north, New Zealand

UNIVERSITÀ DEGLI STUDI
 DI PERUGIA



Control
 ACTH
 °C



Maximum eye temperature for control (■, n=8), local anaesthetic control (▲, n=8), disbudded with local anaesthetic (□, n= 8) and disbudded without local anaesthetic (●, n=6).

Maximum eye temperature following administration (0 min) of ACTH (●, n=6) or saline-control (○, n=6).

Short communication

Evaluation of naturally ventilated dairy barn management by a thermographic method



FLIR T-Series Thermal Imaging camera

Ivana Knížková^{a,*}, Petr Kunc^a, Marie Koubková^b, Jan Flusser^c, Oldřich Dolezal^a

Table 1

Changes of body surface temperature of dairy cows in relation to changes of microclimatic factors (March), mean and S.D. ($N = 12$)

	Cycles	T (°C)	RH (%)	V ($\text{m} \cdot \text{s}^{-1}$)	CWC ($\text{W} \cdot \text{m}^{-2}$)	FP (°C)	B (°C)	HP (°C)
1st day	Closed	16.6	48.2	0.08	195	31.02±1.22	30.22±0.97	30.33±0.80
	Open	13.5	53.1	0.12	462	30.90±1.20	30.30±1.21	30.43±0.72
Difference		-3.1	+5.1	+0.04	+267	-0.12	+0.10	+0.10
2nd day	Open	7.6	60	0.92	965	24.55±1.06	24.14±1.35	24.03±1.06
	Closed	10.0	61	0.17	383	27.52±1.23	27.07±1.43	27.51±1.01
Difference		2.4	+1	0.75	-582	+2.97**	+2.93**	+3.48**
	Closed	10.0	61	0.17	383	27.52±1.23	27.07±1.43	27.51±1.01
	Open	8.5	54	0.84	965	24.11±1.09	23.83±1.57	23.61±1.32
Difference		-1.5	-7	+0.67	+582	-3.41**	-3.24**	-3.90**

**, $P < 0.01$. T , air temperature; RH, relative humidity; V , air velocity; CWC, Canadian wind chill; FR, fore part; B, barrel; HP, hind part.

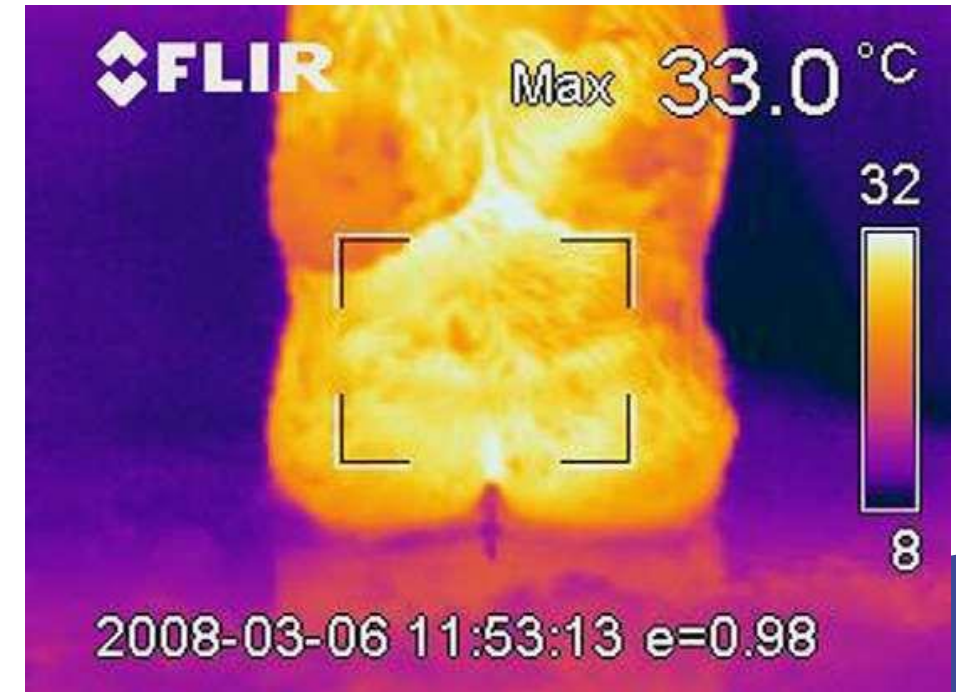
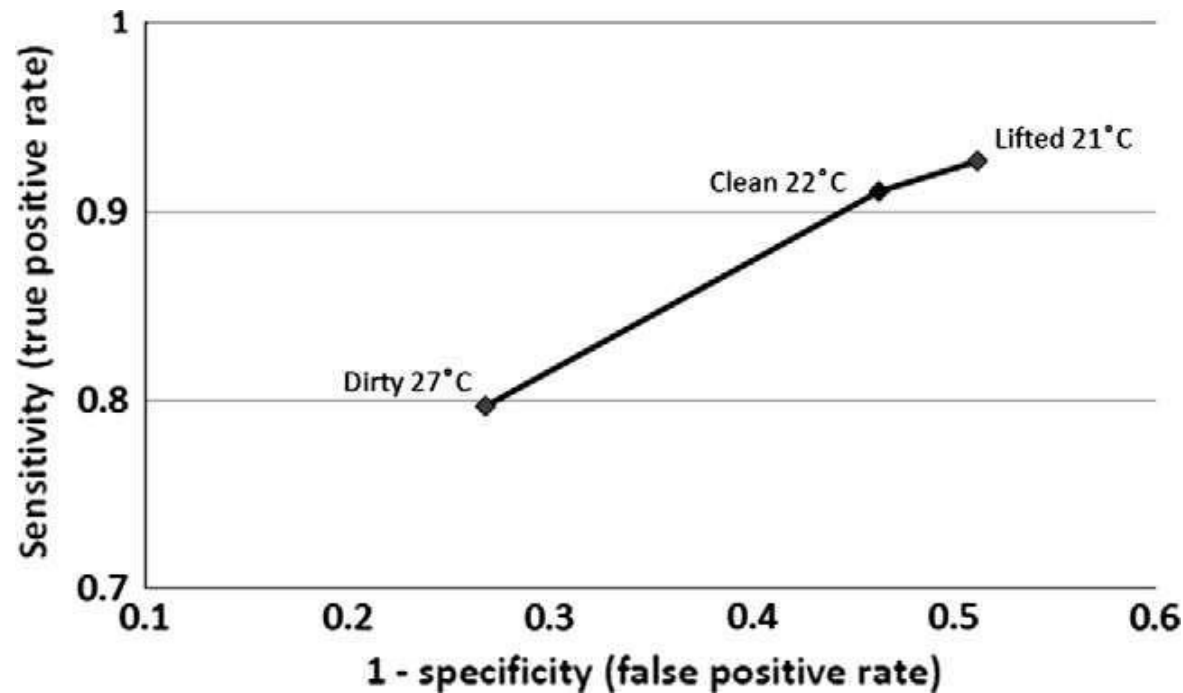
Stokes, J. E. et al (2012): An investigation into the use of infrared thermography (IRT) as a rapid diagnostic tool **for foot lesions** in dairy cattle. *The Veterinary Journal* 193 (3)

ThermaCAM[®]
E2



The ThermaCAM[®] E2 is the smallest, smartest infrared camera ever built.

FLIR
SYSTEMS

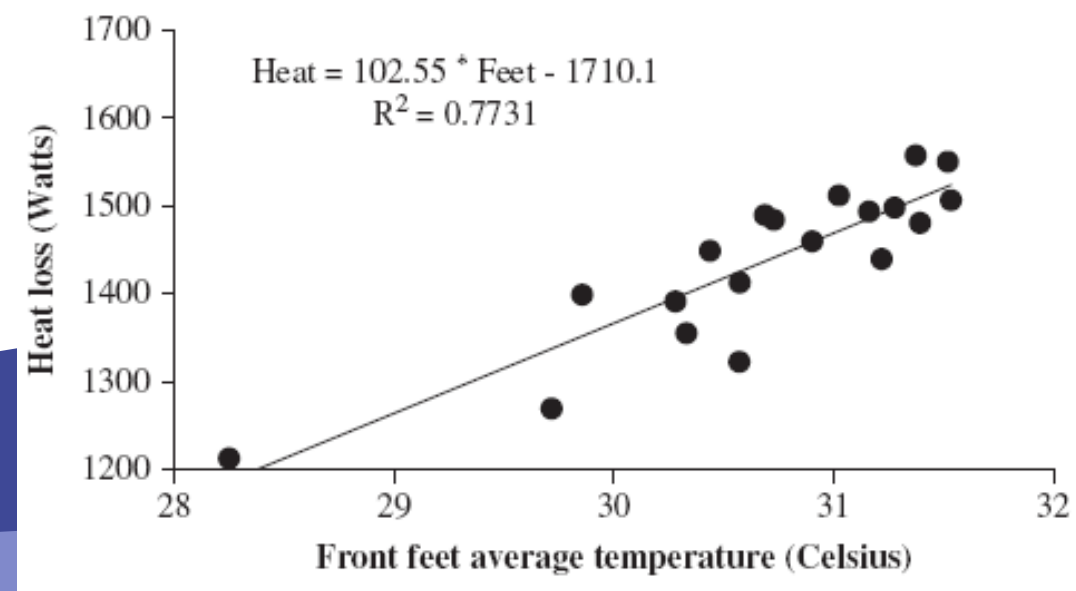
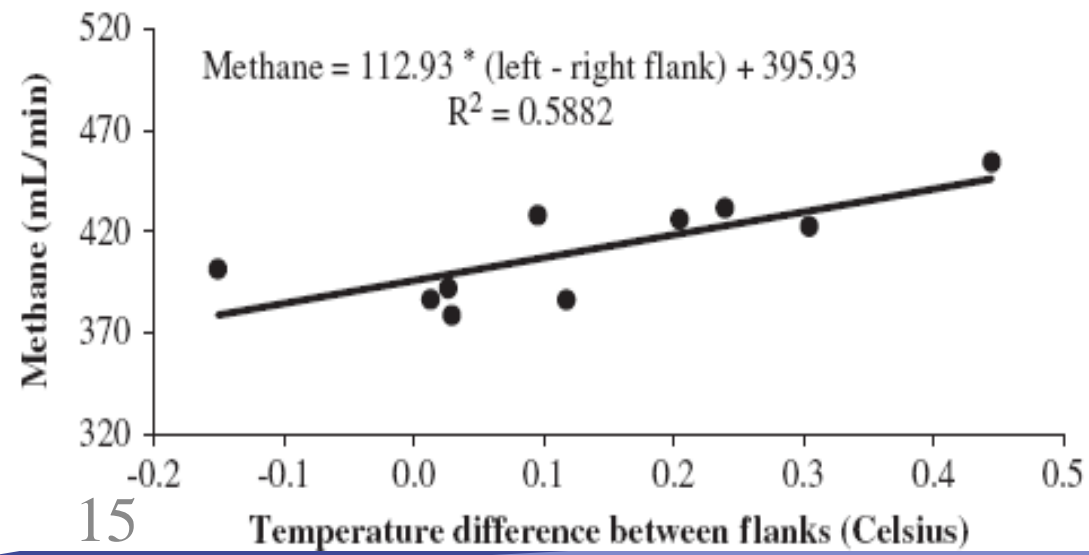
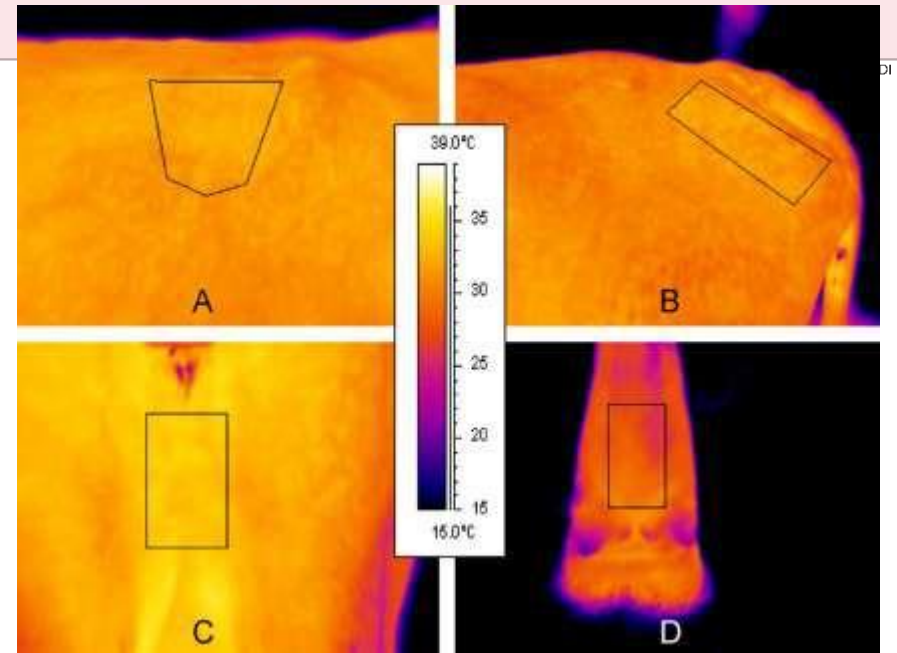


Conclusions: This study established the potential of IRT as a reliable, practical tool for detection of lesions, without having to clean or lift feet but could not be used to differentiate between lesions.

Montanholi, Y. et. al. (2008) Application of infrared thermography as an **indicator of heat and methane production** and its use in the study of skin temperature in response to physiological events in dairy cattle (*Bos taurus*). *Journal of Thermal Biology* 33 (8), S. 468–475.

Pearson correlations between infrared traits and heat production and methane production

Infrared traits	Heat production (P value)	Methane production (P value)
Left flank	0.62 (0.0058) ^a	0.03 (0.8857)
Right flank	0.72 (0.0008) ^a	-0.21 (0.3784)
Left rump	0.71 (0.0010) ^a	-0.01 (0.9607)
Rear area	0.58 (0.0119) ^a	0.03 (0.9155)
Left front foot	0.83 (0.0001) ^a	-0.15 (0.5189)
Right front foot	0.88 (0.0001) ^a	-0.32 (0.1657)
Whole body	0.66 (0.0030) ^a	-0.01 (0.9569)
Trunk	0.66 (0.0028) ^a	-0.05 (0.8235)
Left minus right flank	0.67 (0.0025) ^a	0.53 (0.0165) ^a

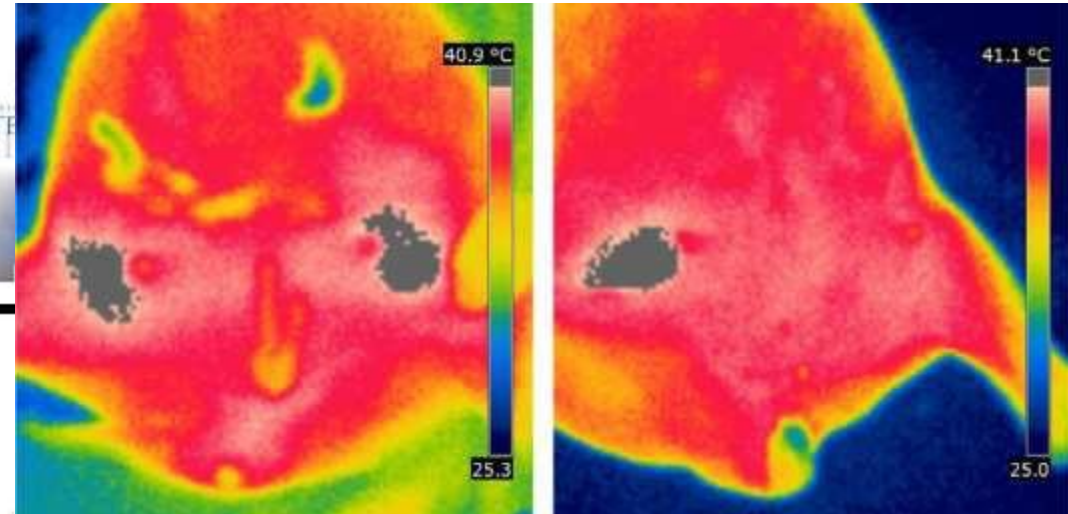




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Mastitis detection in sheep by infrared thermography

Rafhael Felipe Saraiva Martins^a, Tiago do Prado Paim^b, Cyntia de Abreu Cardoso^a,
Bruno Stéfano Lima Dallago^a, Cristiano Barros de Melo^a, Helder Louvandini^b, Concepta McManus^{c,*}

^a Faculty of Agronomy and Veterinary Medicine, University of Brasília, CP 04508, CEP 70910-900 Brasília, Distrito Federal, Brazil

Thermographic images of the udder of ewes in this study, showing an udder with mastitis at both sides (left) and an udder with mastitis at one side (right).

Superficial udder temperatures and milk components from ewes classified as Health, Subclinical and Clinical mastitis.

		Healthy	Subclinical mastitis	Clinical mastitis
Total udder (°C)	Max	38.56 ^b	39.02 ^a	38.4 ^b
	Min	33.56 ^b	33.79 ^a	33.35 ^b
	Mean	36.06 ^{ns}	36.3 ^{ns}	35.89 ^{ns}
Left side (°C)	Front	42.21 ^{ns}	37.33 ^{ns}	36.45 ^{ns}
	Intermediate	37.19 ^b	37.50 ^a	36.90 ^b
	Rear	37.17 ^a	37.42 ^b	36.69 ^c
	Mean	37.01 ^b	37.48 ^a	36.60 ^b
Right side (°C)	Front	36.85 ^{b,a}	37.14 ^a	36.53 ^b
	Intermediate	37.22 ^b	37.72 ^a	36.84 ^b
	Rear	37.16 ^a	37.50 ^a	36.71 ^b
	Mean	37.08 ^b	37.44 ^a	36.74 ^b
TCL (%)	Fat	5.68 ^b	6.17 ^b	7.12 ^a
	Protein	5.40 ^b	5.17 ^b	5.82 ^a
	Lactose	5.00 ^a	4.91 ^a	4.60 ^b
	FFDM	11.25 ^a	10.99 ^b	11.29 ^a
	TDM	17.43 ^b	16.67 ^c	18.41 ^a
SCC (×1000 cells/mL)		167.1 ^b	540.7 ^b	2693.2 ^a

Conclusions: The results demonstrate that infrared udder temperatures can be a good auxiliary diagnostic method to mastitis in sheep, principally to subclinical mastitis.



Correlations between eye temperature and performance breeding values.

Trait	Breeding value of temperature traits					
	ET_B	ET_JA	ET_A	ET_BJA	ET_JAA	
Breeding value of performance traits	Walk Score	-0.00	0.07	0.07	-0.10	-0.13
	Trot Score	0.02	0.19*	0.12	0.00	-0.05
	Canter Score	0.07	0.22*	0.14	-0.04	0.02
	Submission Score	0.00	0.22*	0.11	-0.03	-0.00
	General Impression Score	0.02	0.28*	0.10	-0.06	0.02
	Total Dressage Score	-0.00	0.21*	0.09	-0.03	-0.03

ET_B = eye temperature taken 3 h before the competition; ET_JA = eye temperature taken just after the competition (<5 min after the dressage exercise) ET_A = eye temperature taken 3 h after the competition, when the animal was resting; ET_BJA = Difference between eye temperature taken 3 h before the competition and eye temperature taken just after the competition (<5 min after the dressage exercise) and ET_JAA = Difference between eye temperature taken just after the competition (<5 min after the dressage exercise) and eye temperature taken 3 h after the competition, when the animal was resting. * $p < 0.05$; ** $p < 0.01$; *** $p < 0.001$.

Authors analyzed different factors influencing stress of horses during Dressage competitions, evaluated with IRT. Correlations with Dressage results were also assessed.

Conclusions: These findings indicate that it is **possible to assess selection for stress with ET in horses.**

The positive medium **correlations** found suggested that **horses with higher ET values were better dressage performers**, appearing to be related with a **more proactive and energetic response** of the horse to the environmental stimuli and hence, to physiological stress rather than emotional stress (or “distress”).

Genetic study of stress assessed with infrared thermography during dressage competitions in the Pura Raza Español horse

María José Sánchez^a, Ester Bartolomé^{b,*}, Mercedes Valera^a

General Linear Model and post-hoc LSM test analysis of the environmental effects in thermography and Dressage performance variables in Pura Raza Español breed

Traits		Stud	Trip	Training	Rider	Age (LSMean)			Event (LSMean)		
						4	5	6	Event 1	Event 2	Event 3
						years	years	years			
Temperature traits	ET_B	***	***	***	***	35.4 ^a	35.2 ^{ab}	35.0 ^a	35.4 ^a	35.5 ^a	34.9 ^b
	ET_JA	***	***	***	***	36.6 ^a	36.4 ^a	35.9 ^b	36.9 ^a	36.6 ^a	35.5 ^b
	ET_A	***	***	***	***	36.4 ^a	35.8 ^b	35.6 ^b	36.2 ^a	36.6 ^a	35.3 ^b
	ET_BJA	***	***	*	***	0.57 ^a	0.44 ^a	0.50 ^a	0.58 ^{ab}	0.64 ^a	0.27 ^b
	ET_JAA	***	***	*	***	-0.24 ^a	-0.58 ^a	-0.32 ^a	-0.74 ^a	-0.10 ^b	-0.12 ^b
Performance traits	Walk Score	***	***	***	***	6.4 ^b	6.7 ^a	6.8 ^a	6.1 ^b	6.7 ^a	6.8 ^a
	Trot Score	***	***	***	***	6.4 ^b	6.7 ^a	6.7 ^{ab}	6.2 ^b	6.8 ^a	6.9 ^a
	Canter Score	***	***	***	***	6.6 ^a	6.7 ^a	6.7 ^a	6.2 ^b	6.9 ^a	6.9 ^a
	Submission Score	***	***	***	***	6.3 ^a	6.5 ^a	6.6 ^a	6.1 ^b	6.7 ^a	6.7 ^a
	General Impression Score	***	***	***	***	6.4 ^a	6.7 ^a	6.6 ^a	6.1 ^b	6.7 ^a	6.8 ^a
	Total Dressage Score	***	***	***	***	63.5 ^b	66.5 ^a	65.9 ^{ab}	60.4 ^b	67.3 ^a	68.2 ^a

ET_B = eye temperature taken 3 h before the competition; ET_JA = eye temperature taken just after the competition (<5 min after the dressage exercise); ET_A = eye temperature taken 3 h after the competition, when the animal was resting; ET_BJA = difference between eye temperature taken 3 h before the competition and eye temperature taken just after the competition (<5 min after the dressage exercise) and ET_JAA = difference between eye temperature taken just after the competition (<5 min after the dressage exercise) and eye temperature taken 3 h after the competition, when the animal was resting. ^{abc}Values within a row with different superscripts differ significantly at * $p < 0.05$; ** $p < 0.01$; *** $p < 0.001$

Mean (and standard deviation) of the marginal posterior distributions means for heritabilities, repeatability and the rider ratio, for all the traits analyzed.

Trait	Heritabilities	Repeatability	Rider ratio	
Temperature traits	ET_B	0.38(0.173)	0.76(0.343)	0.45(0.127)
	ET_JA	h² = 14% to 50%	0.25(0.153)	
	ET_A		0.77(0.431)	
	ET_BJA		0.30(0.204)	
	ET_JAA		0.31(0.180)	
Performance traits	Walk Score		0.44(0.183)	0.79(0.370)
	Trot Score	0.40(0.187)	0.76(0.370)	0.22(0.117)
	Canter Score	0.43(0.176)	0.78(0.352)	0.20(0.113)
	Submission Score	0.37(0.163)	0.71(0.326)	0.28(0.123)
	General Impression Score	0.40(0.168)	0.73(0.335)	0.26(0.127)
	Total Dressage Score	0.42(0.179)	0.73(0.350)	0.24(0.121)

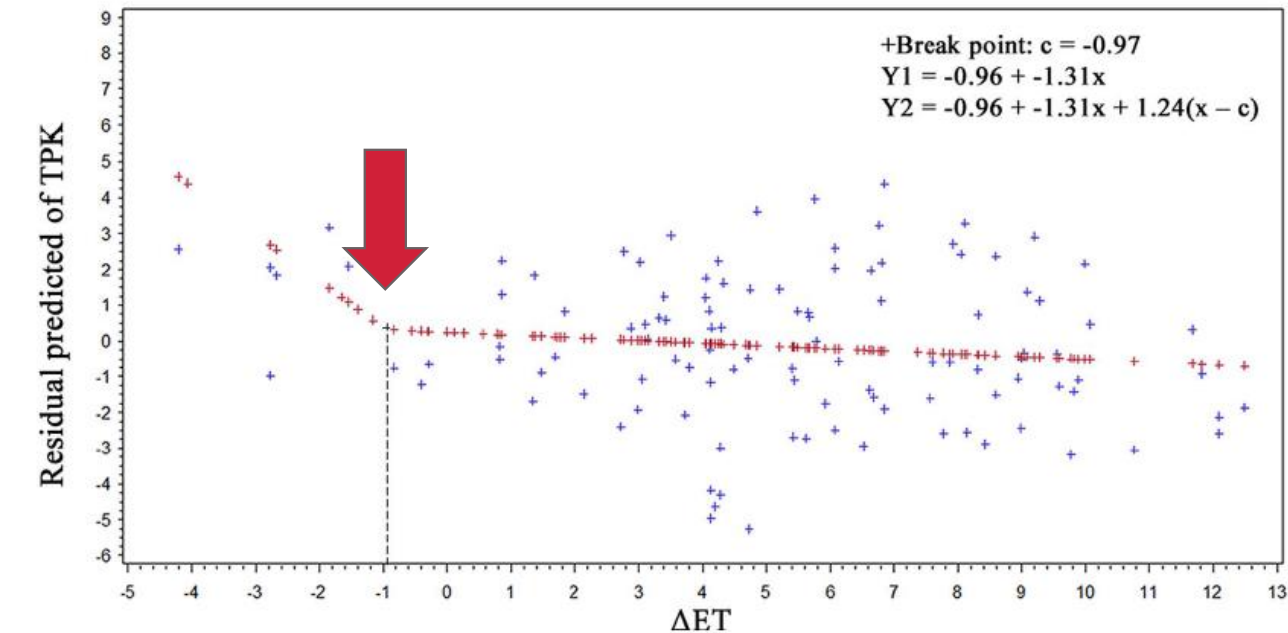
ET_B = eye temperature taken 3 h before the competition; ET_JA = eye temperature taken just after the competition (<5 min after the dressage exercise); ET_A = eye temperature taken 3 h after the competition, when the animal was resting; ET_BJA = difference between eye temperature taken 3 h before the competition and eye temperature taken just after the competition (<5 min after the dressage exercise) and ET_JAA = difference between eye temperature taken just after the competition (<5 min after the dressage exercise) and eye temperature taken 3 h after the competition, when the animal was resting.

Stress level effects on sport performance during trotting races in Spanish Trotter Horses

Negro S, Bartolomé E, Molina A et al. See more

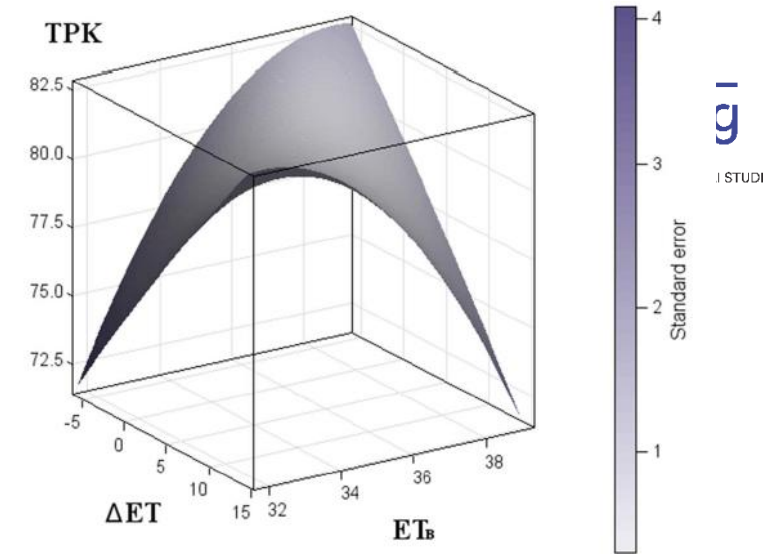
Research in Veterinary Science (2018) 118 86-90

DOI: 10.1016/j.rvsc.2018.01.017



Segmented regression analysis and break point for the residual predicted of time per kilometre (TPK) according to the eye temperature increase (ΔET) in the Spanish Trotter Horse participating in trotting races.

Authors analyzed the stress of Trotter Horses during Trotting Races with IRT and Heart Rate. Developed a regression analysis and a response surface comparing results (TPK) and stress (ET increase and ET before competition).



Conclusions: Results suggest that racing performance is influenced by the level of physiological stress, because ET, but not HR, are related to competition results. This study also suggests an **elliptical behaviour of ET during the race which would be incremental to a certain point from which the animals would suffer the consequences of distress, diminishing its performance and worsening its racing results.**



Stress at rest in working dogs assessed with infrared thermography

E. Bartolomé*, D.I. Perdomo-González, M.J. Sánchez-Guerrero, M. Valera

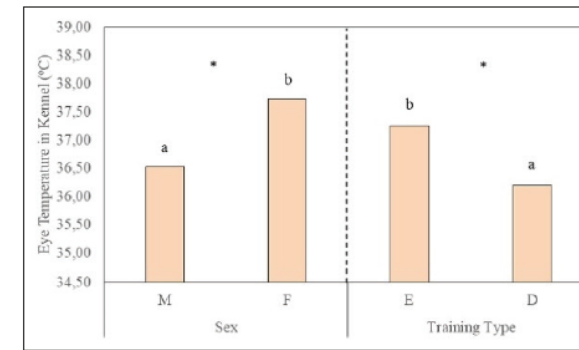
Table 1. Pearson's correlations between eye temperature variables (measured in dog kennels -ETK-, inside the police cars -ETC-, their difference -ETD- and mean value -MET-, including a thermographic photo) and heart rate -HR- variable (measured inside the police car) analyzed. Mean (absolute values), standard deviation and coefficient of variation (in percentage in parentheses), appear in the diagonal.

	ETK	ETC	ETD	HR	MET
					
ETK	36.79±1.09 (2.98%)	0.38 ^{n.s.}	0.61 ^{**}	0.23 ^{n.s.}	0.85 ^{***}
ETC		36.14±1.00 (2.78%)	0.50 [*]	- 0.14 ^{n.s.}	0.82 ^{***}
ETD			0.92±0.95 (103.04%)	0.34 ^{n.s.}	0.09 ^{n.s.}
HR				92.33±18.54 (20.08%)	0.06 ^{n.s.}
MET					36.47±0.87 (2.39%)

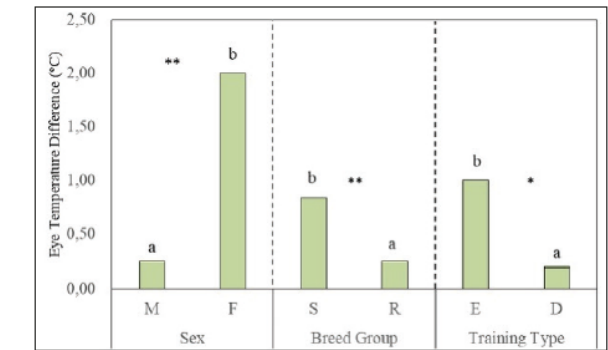
Where ETD= ETC – ETK; MET = (ETC + ETK)/2; *p<0.05; **p<0.01; *** p<0.001; n.s. not statistically significant.

Figure 2. General Lineal Model and Duncan post-hoc test for variables that resulted statistically significant (eye temperature assessed in kennel and eye temperature difference between kennel and police car) for sex, breed group and training type effects.

2A. Eye Temperature in Kennel



2B. Eye Temperature Difference



Where M = male; F = female; S = Shepherd dogs; R = Retriever dogs; E = explosives; D = drugs; *p<0.05; **p<0.01; *** p<0.001. Different letters indicated statistically significant differences between means (p<0.05).

Authors analyzed the stress of Police working dogs trained for drugs and explosives detection at rest, evaluated on their kennels and on the car they used for working purposes. ET and HR were assessed.

Conclusions: Statistically significant differences were reported between stress perceived in the kennel versus stress perceived in the police car. The stress of working dogs shown in the kennel and the size of the stress difference recorded between the kennel and the police car was influenced by environmental factors such as sex, breed group or training type.



Youtube video: "Infrared Thermography for Veterinary" – Pass Thermal (3:00min.)

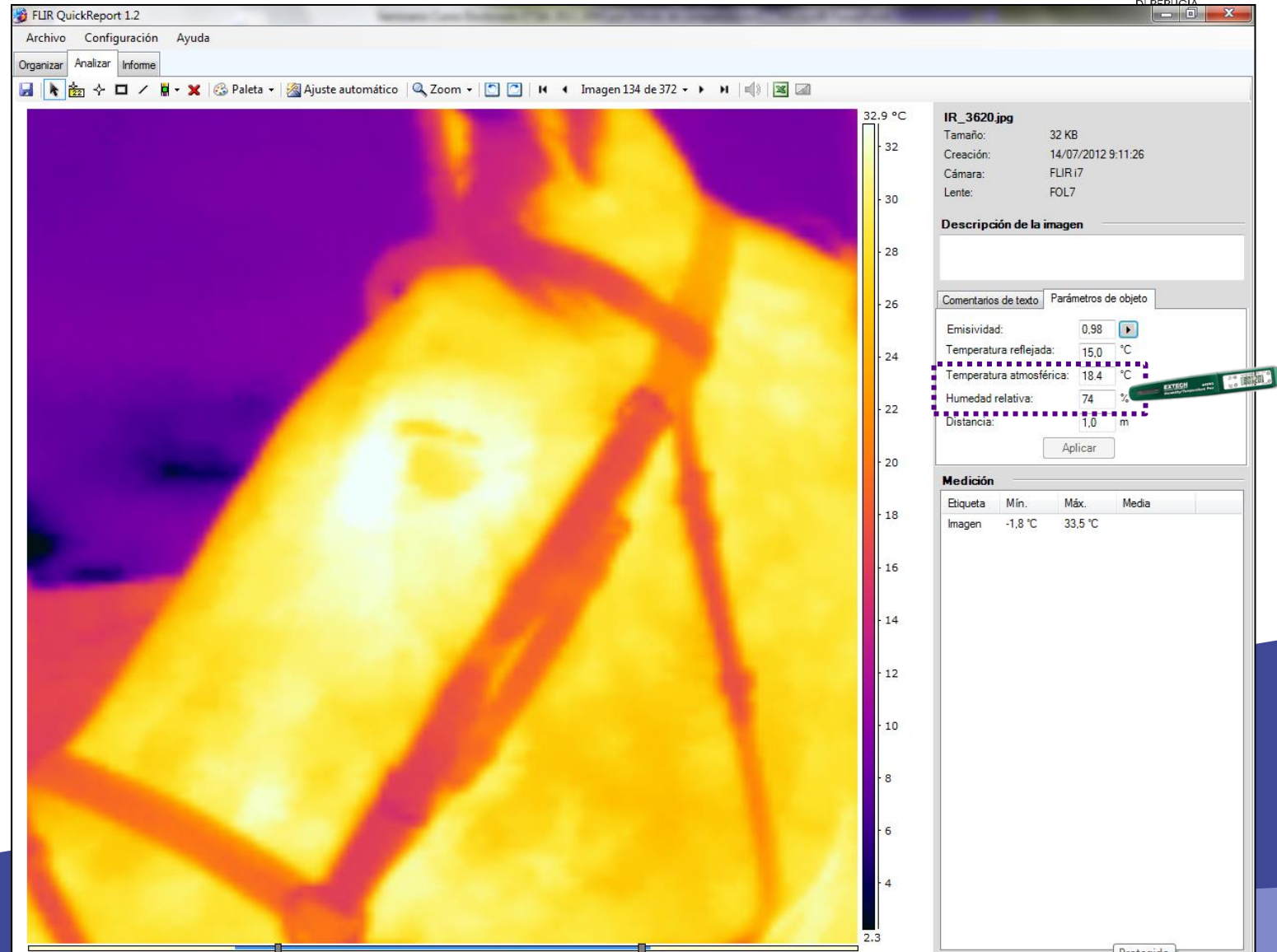
Índex

1. What is Infrared Thermography?
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HOW DO WE OBTAIN THE DATA

IRT camera assigns color to the infrared radiation:
different color equals different temperature.

IRT data has to be **analyzed** with an **external software**, where *Emissivity*, *Environmental Temperature* and *Relative Humidity* and *Area* of the image to be measured, have to be settled.

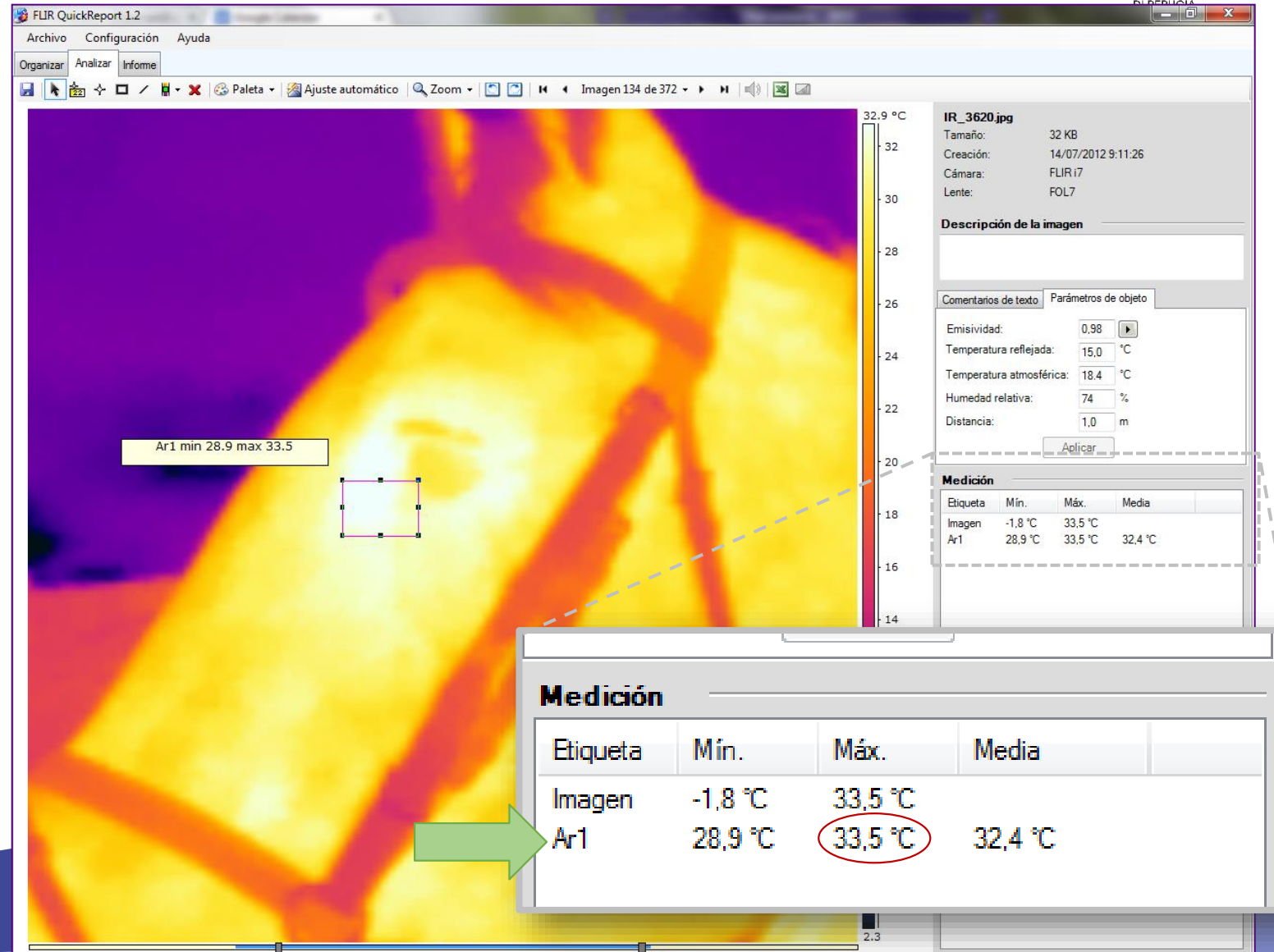


How do we use it?

HOW DO WE OBTAIN THE DATA

IRT camera assigns color to the infrared radiation:
different color equals different temperature.

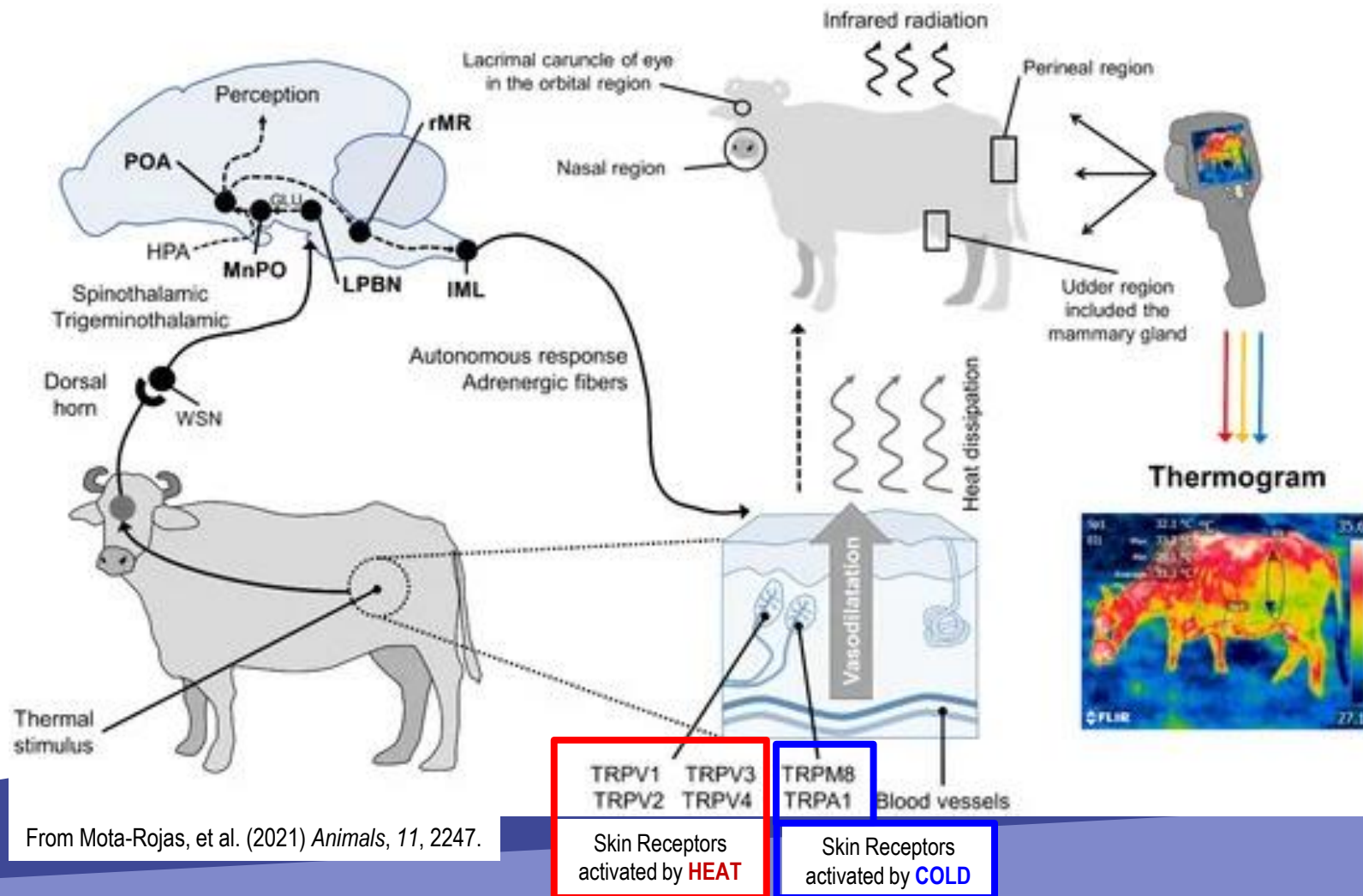
IRT data has to be analyzed with an **external software**, where *Emissivity*, *Environmental Temperature* and *Relative Humidity* and *Area* of the image to be measured, have to be settled.



WHAT ARE WE MEASURING?

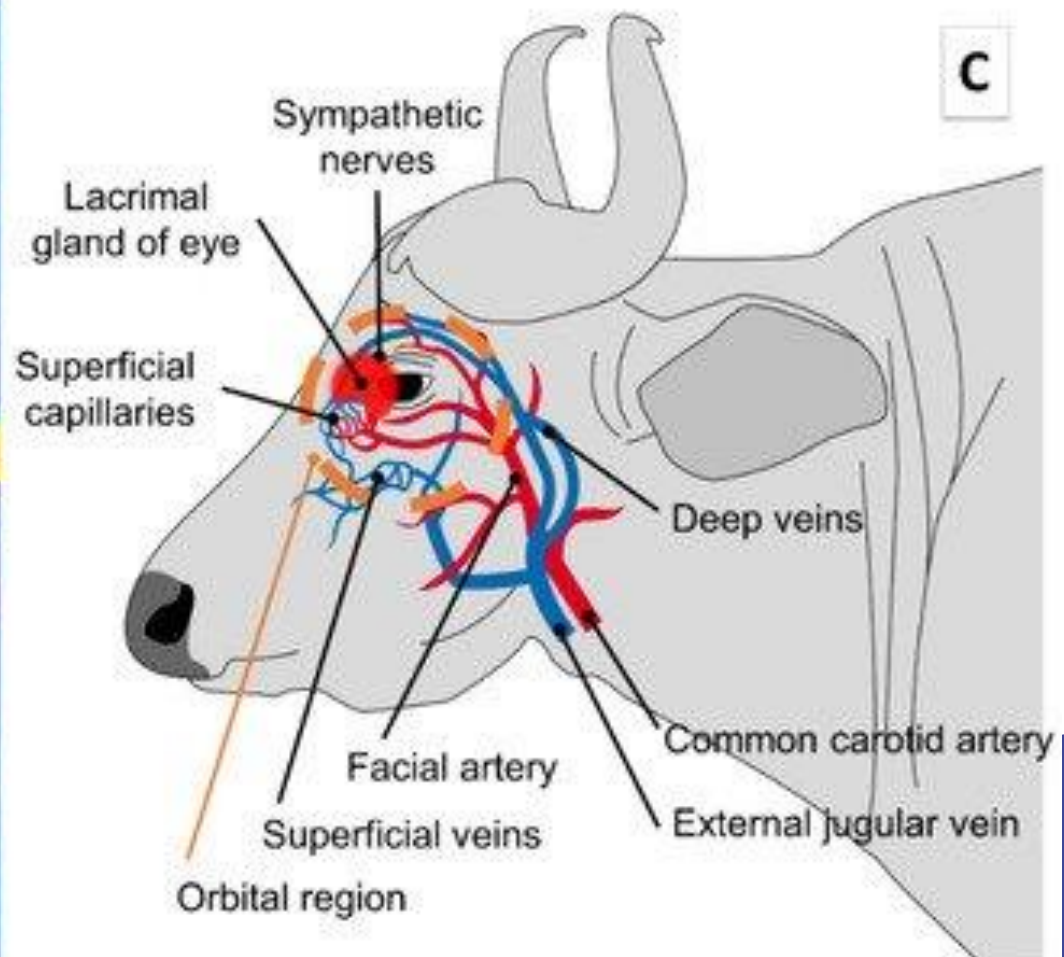
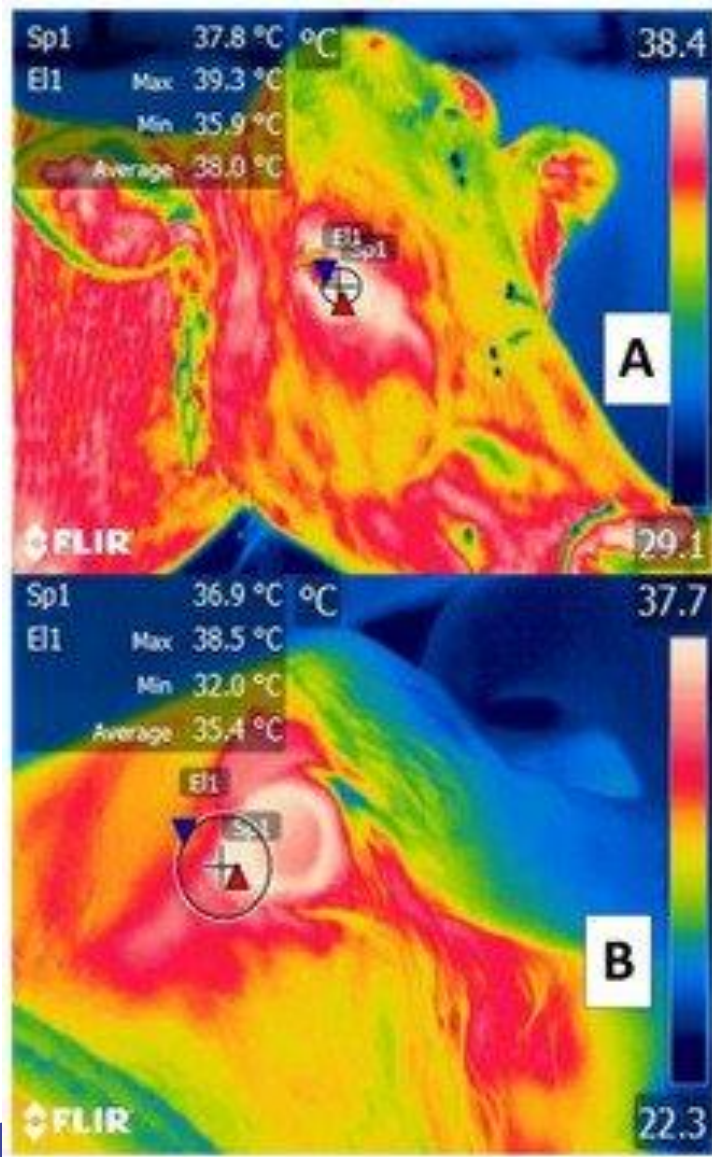
Hypothalamic neuromodulation of thermoregulation and its cutaneous response.

- ✓ The thermoregulatory response **begins** on with **thermoreceptors in the dermis**.
- ✓ These **receive afferent information** from a thermal stimulus and **send the signal** to the laminae of the dorsal horn of the **spinal cord**.
- ✓ The **heat-sensitive (WSN) spinothalamic and trigeminothalamic neurons** in this zone **relay the impulse** to third-order neurons in brain structures.
- ✓ From there, **they are projected to the median preoptic nucleus (MnPO) in the preoptic area (POA) of the hypothalamus**.
- ✓ The hypothalamic network is responsible for **integrating behavioral, neuroendocrine** (mediated by the hypothalamic–pituitary–adrenal axis (HPA)), and **autonomic thermoregulatory effector responses**.

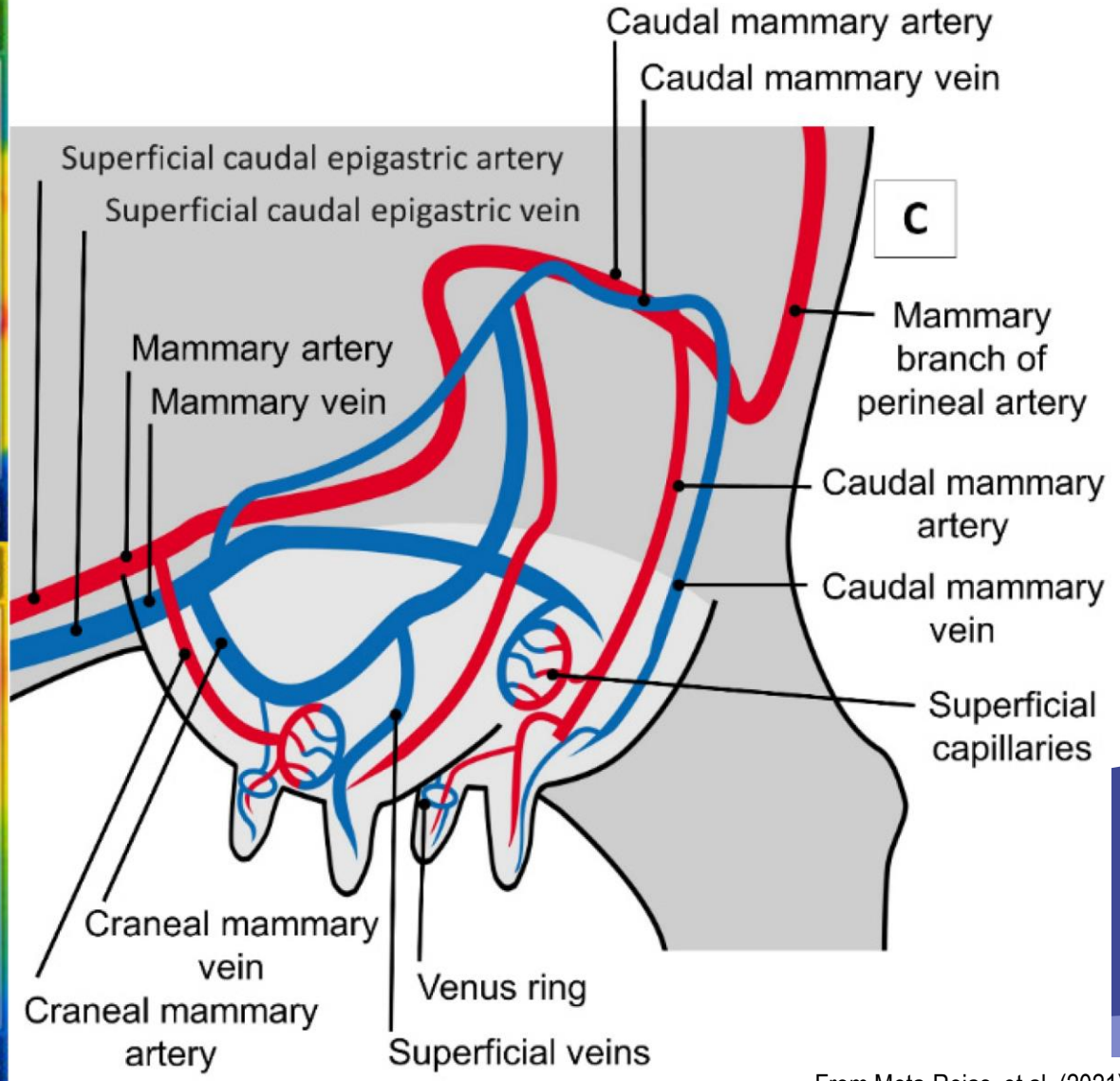
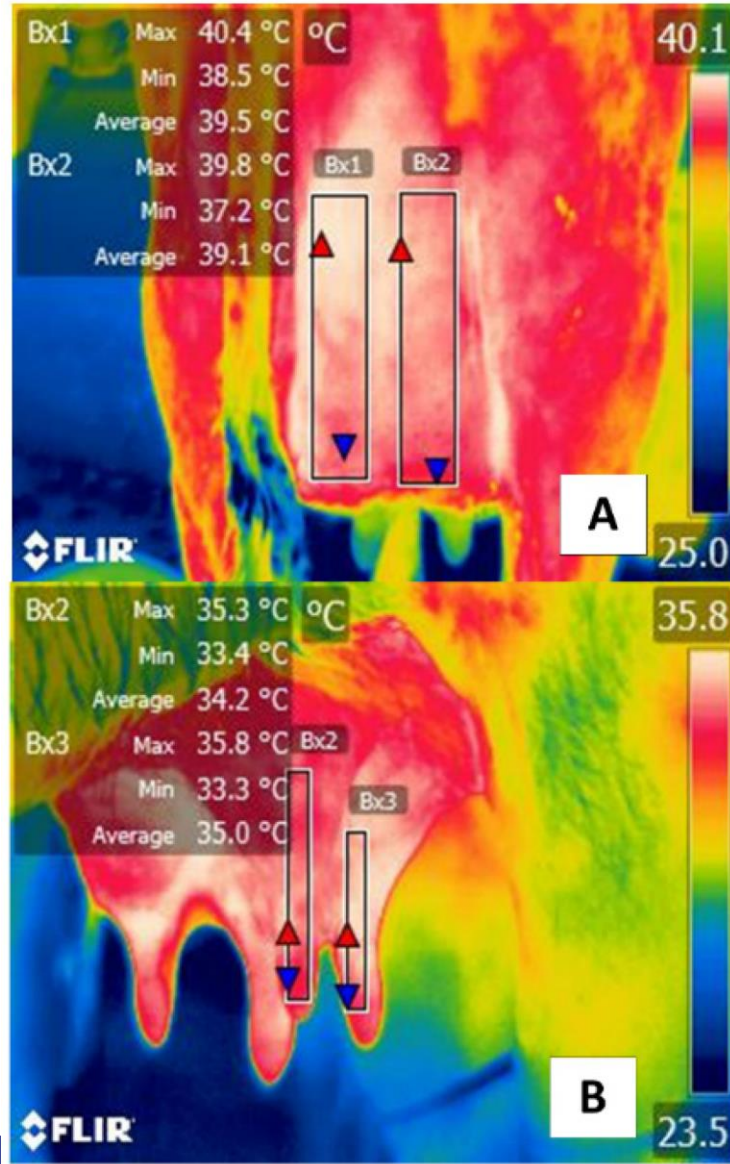


How do we use it?

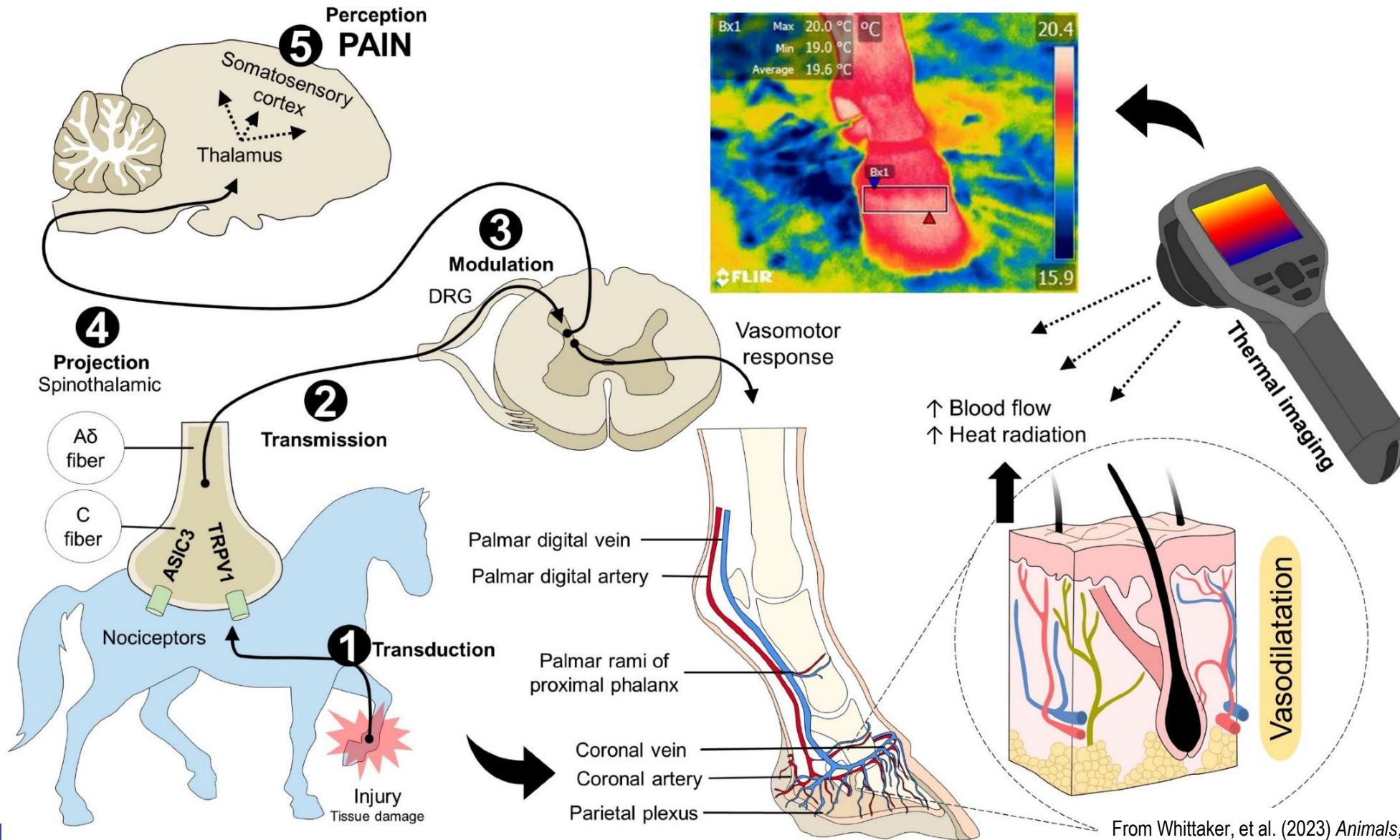
WHAT ARE WE MEASURING?



WHAT ARE WE MEASURING?



WHAT ARE WE MEASURING?



From Whittaker, et al. (2023) *Animals*, 13(13), 2065.

Índex

1. What is Infrared Thermography?
2. Applications in Animal Production.
3. How do we use it?
- 4. Practical exercises.**

SETOUT

- We are going to divide the big group in **4 working groups (purple, blue, yellow, orange)**, with 4-5 students per group.
- Each group is going to **analyze a different practical case** from already **published papers**.
- Each group **have to solve** the following **points** for their case:
 - I. Species and Production System.*
 - II. Problem/Issue detected/studied on the animal/s.*
 - III. Proposal to prevent this problem and to collect information regularly.*
- Finally, **one student from each group** will come out and **present their proposal** to the rest of the **students that will give their feed-back** about it.



PURPLE GROUP



Figure from: Zheng, S.; Zhou, C.; Jiang, X.; Huang, J.; Xu, D. Progress on Infrared Imaging Technology in Animal Production: A Review. *Sensors* **2022**, *22*, 705. <https://doi.org/10.3390/s22030705>



BLUE GROUP

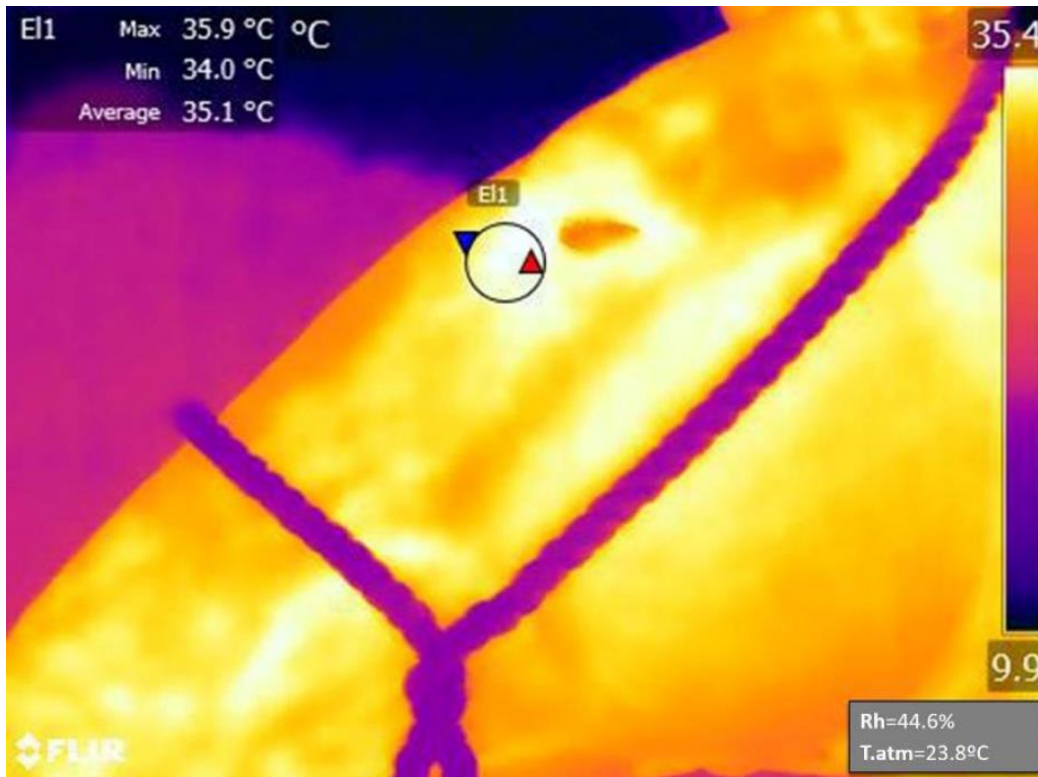


Figure from: Bartolomé, E.; Perdomo-González, D.I.; Ripollés-Lobo, M.; Valera, M. Basal Reactivity Evaluated by Infrared Thermography in the “Caballo de Deporte Español” Horse Breed According to Its Coat Color. *Animals* **2022**, *12*, 2515. <https://doi.org/10.3390/ani12192515>



YELLOW GROUP

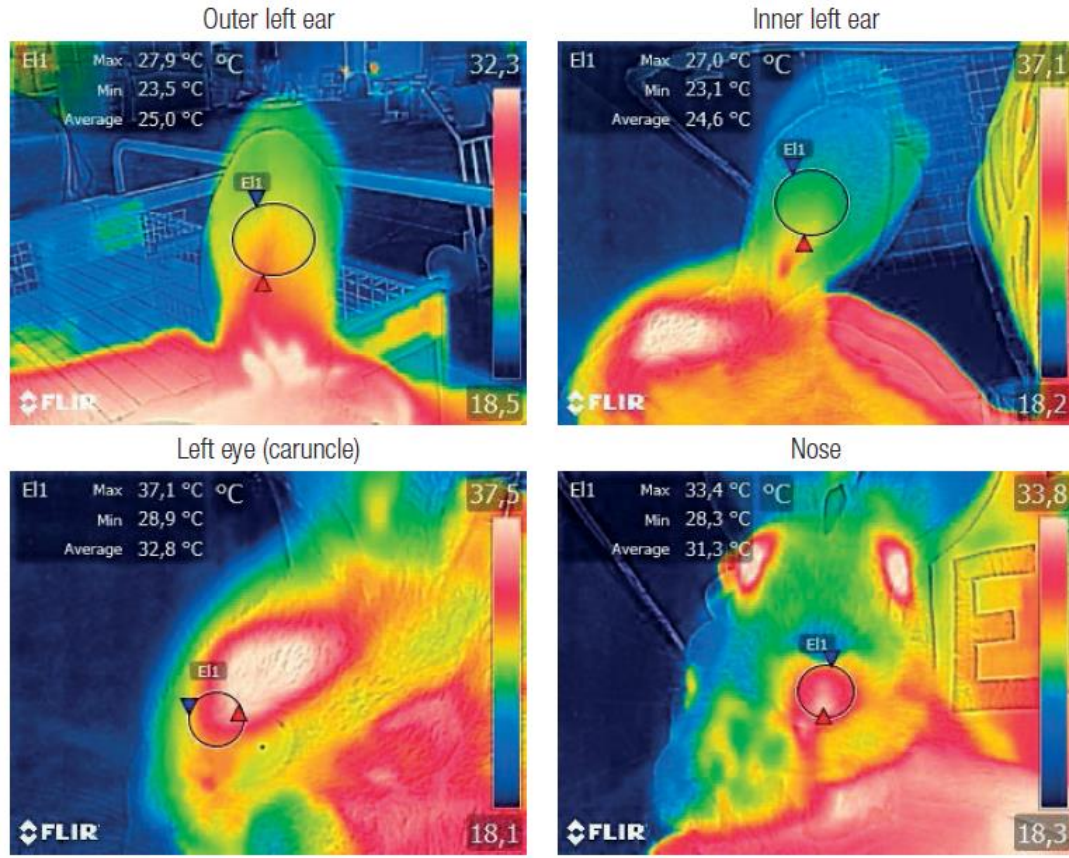


Figure from: Jaén-Téllez J.A., Bartolomé E., Sánchez-Guerrero M.J., Valera M., González-Redondo (2021) Relationship between rectal temperature measured with a conventional thermometer and the temperature of several body regions measured by infrared thermography in fattening rabbits. Influence of different environmental factors. *World Rabbit Science*, 29 (4), pp. 263 – 273. DOI: 10.4995/wrs.2021.15556.



ORANGE GROUP

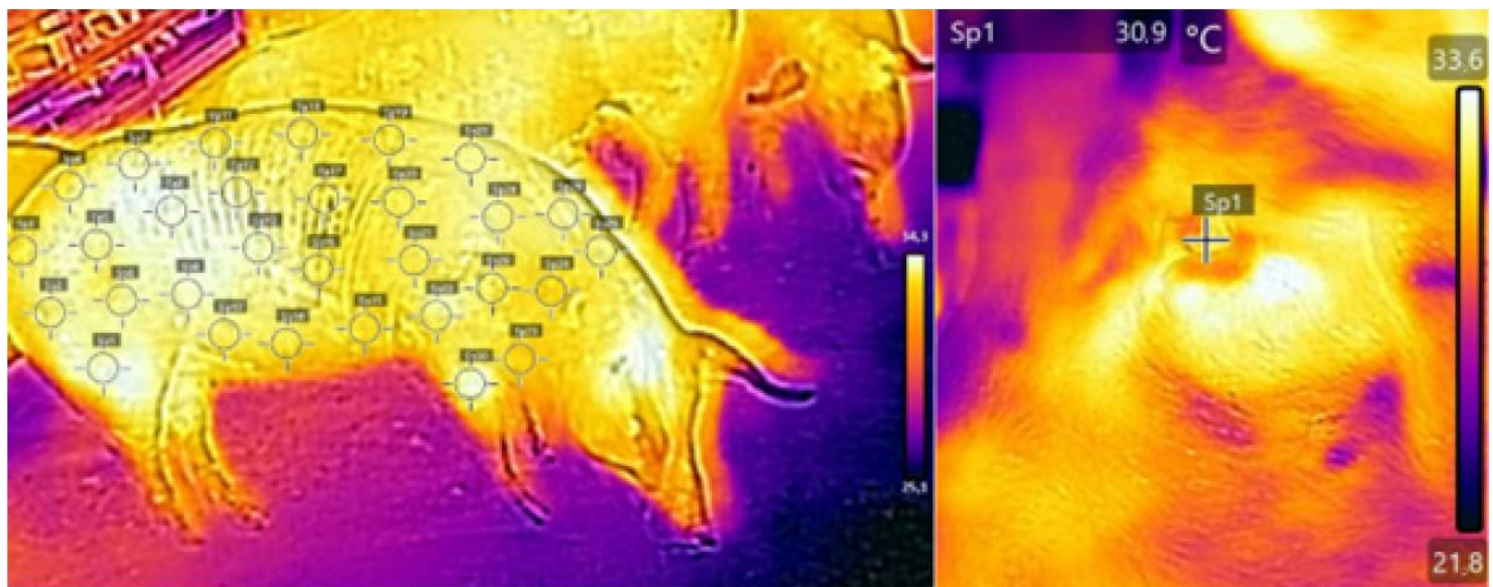


Figure from: Palermo Mendes, J.; Ribeiro Caldara, F.; de Castro Burbarelli, M.F.; Valentim, J.K.; Ferreira de Brito Mandú, D.; Garófallo Garcia, R.; Correia de Lima Almeida-Paz, I.; Markiy Odakura, A.; Lourenço da Silva, M.I. Performance and Welfare of Sows Exposed to Auditory Environmental Enrichment in Mixed or Collective Housing Systems. *Animals* **2023**, *13*, 1226. <https://doi.org/10.3390/ani13071226>.



Thank you

